

Journal of Parasitology and Vector Biology

Volume 5 Number 4 April 2013
ISSN 2141-2510



*Academic
Journals*

ABOUT JPVB

The **Journal of Parasitology and Vector Biology (JPVB)** is published monthly (one volume per year) by Academic Journals.

Journal of Parasitology and Vector Biology (JPVB) provides rapid publication (monthly) of articles in all areas of the subject such as Parasitism, Helminthology, Cloning vector, retroviral integration, Genetic markers etc.

Submission of Manuscript

Submit manuscripts as e-mail attachment to the Editorial Office at: jpvb@academicjournals.org.
A manuscript number will be mailed to the corresponding author shortly after submission.

For all other correspondence that cannot be sent by e-mail, please contact the editorial office (at jpvb@academicjournals.org).

The Journal of Parasitology and Vector Biology will only accept manuscripts submitted as e-mail attachments.

Please read the **Instructions for Authors** before submitting your manuscript. The manuscript files should be given the last name of the first author.

Editors

Dr. Ratna Chakrabarti

*Department of Molecular Biology and Microbiology,
University of Central Florida,
Biomolecular Research Annex,
12722 Research Parkway,
Orlando,
USA.*

Dr. Rajni Kant

*Scientist D (ADG),
(P&I Division) Indian Council of Medical Research
Post Box 4911, Ansari Nagar,
New Delhi-110029
India.*

Dr. Ramasamy Harikrishnan

*Faculty of Marine Science, College of Ocean
Sciences
Jeju National University
Jeju city, Jeju 690 756
South Korea.*

Dr. Rokkam Madhavi

*Andhra University
Visakhapatnam - 530003
Andhra Pradesh
India.*

Dr. Mukabana Wolfgang Richard

*School of Biological Sciences
University of Nairobi
P.O. Box 30197 - 00100 GPO
Nairobi,
Kenya.*

Dr. Lachhman Das Singla

*College of Veterinary Science
Guru Angad Dev Veterinary and Animal Sciences
University
Ludhiana-141004
Punjab
India.*

Editorial Board

Dr. Imna Issa Malele

*Tsetse & Trypanosomiasis Research Institute
Tanzania.*

Dr. Mausumi Bharadwaj

*Institute of Cytology & Preventive Oncology,
(Indian Council of Medical Research)
I-7, Sector - 39
Post Box No. 544
Noida - 201 301
India.*

Dr. James Culvin Morris

*Clemson University
214 Biosystems Research Complex
Clemson SC 29634
USA.*

Instructions for Author

Electronic submission of manuscripts is strongly encouraged, provided that the text, tables, and figures are included in a single Microsoft Word file (preferably in Arial font).

The **cover letter** should include the corresponding author's full address and telephone/fax numbers and should be in an e-mail message sent to the Editor, with the file, whose name should begin with the first author's surname, as an attachment.

Article Types

Three types of manuscripts may be submitted:

Regular articles: These should describe new and carefully confirmed findings, and experimental procedures should be given in sufficient detail for others to verify the work. The length of a full paper should be the minimum required to describe and interpret the work clearly.

Short Communications: A Short Communication is suitable for recording the results of complete small investigations or giving details of new models or hypotheses, innovative methods, techniques or apparatus. The style of main sections need not conform to that of full-length papers. Short communications are 2 to 4 printed pages (about 6 to 12 manuscript pages) in length.

Reviews: Submissions of reviews and perspectives covering topics of current interest are welcome and encouraged. Reviews should be concise and no longer than 4-6 printed pages (about 12 to 18 manuscript pages). Reviews are also peer-reviewed.

Review Process

All manuscripts are reviewed by an editor and members of the Editorial Board or qualified outside reviewers. Authors cannot nominate reviewers. Only reviewers randomly selected from our database with specialization in the subject area will be contacted to evaluate the manuscripts. The process will be blind review.

Decisions will be made as rapidly as possible, and the journal strives to return reviewers' comments to authors as fast as possible. The editorial board will re-review manuscripts that are accepted pending revision. It is the goal of the JPVB to publish manuscripts within weeks after submission.

Regular articles

All portions of the manuscript must be typed double-spaced and all pages numbered starting from the title page.

The Title should be a brief phrase describing the contents of the paper. The Title Page should include the authors' full names and affiliations, the name of the corresponding author along with phone, fax and E-mail information. Present addresses of authors should appear as a footnote.

The Abstract should be informative and completely self-explanatory, briefly present the topic, state the scope of the experiments, indicate significant data, and point out major findings and conclusions. The Abstract should be 100 to 200 words in length. Complete sentences, active verbs, and the third person should be used, and the abstract should be written in the past tense. Standard nomenclature should be used and abbreviations should be avoided. No literature should be cited. Following the abstract, about 3 to 10 key words that will provide indexing references should be listed.

A list of non-standard **Abbreviations** should be added. In general, non-standard abbreviations should be used only when the full term is very long and used often. Each abbreviation should be spelled out and introduced in parentheses the first time it is used in the text. Only recommended SI units should be used. Authors should use the solidus presentation (mg/ml). Standard abbreviations (such as ATP and DNA) need not be defined.

The Introduction should provide a clear statement of the problem, the relevant literature on the subject, and the proposed approach or solution. It should be understandable to colleagues from a broad range of scientific disciplines.

Materials and methods should be complete enough to allow experiments to be reproduced. However, only truly new procedures should be described in detail; previously published procedures should be cited, and important modifications of published procedures should be mentioned briefly. Capitalize trade names and include the manufacturer's name and address. Subheadings should be used. Methods in general use need not be described in detail.

Results should be presented with clarity and precision.

The results should be written in the past tense when describing findings in the authors' experiments. Previously published findings should be written in the present tense. Results should be explained, but largely without referring to the literature. Discussion, speculation and detailed interpretation of data should not be included in the Results but should be put into the Discussion section.

The Discussion should interpret the findings in view of the results obtained in this and in past studies on this topic. State the conclusions in a few sentences at the end of the paper. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined.

The Acknowledgments of people, grants, funds, etc should be brief.

Tables should be kept to a minimum and be designed to be as simple as possible. Tables are to be typed double-spaced throughout, including headings and footnotes. Each table should be on a separate page, numbered consecutively in Arabic numerals and supplied with a heading and a legend. Tables should be self-explanatory without reference to the text. The details of the methods used in the experiments should preferably be described in the legend instead of in the text. The same data should not be presented in both table and graph form or repeated in the text.

Figure legends should be typed in numerical order on a separate sheet. Graphics should be prepared using applications capable of generating high resolution GIF, TIFF, JPEG or Powerpoint before pasting in the Microsoft Word manuscript file. Tables should be prepared in Microsoft Word. Use Arabic numerals to designate figures and upper case letters for their parts (Figure 1). Begin each legend with a title and include sufficient description so that the figure is understandable without reading the text of the manuscript. Information given in legends should not be repeated in the text.

References: In the text, a reference identified by means of an author's name should be followed by the date of the reference in parentheses. When there are more than two authors, only the first author's name should be mentioned, followed by 'et al'. In the event that an author cited has had two or more works published during the same year, the reference, both in the text and in the reference list, should be identified by a lower case letter like 'a' and 'b' after the date to distinguish the works.

Examples:

Abayomi (2000), Agindotan et al. (2003), (Kelebeni, 1983), (Usman and Smith, 1992), (Chege, 1998;

1987a,b; Tijani, 1993,1995), (Kumasi et al., 2001)

References should be listed at the end of the paper in alphabetical order. Articles in preparation or articles submitted for publication, unpublished observations, personal communications, etc. should not be included in the reference list but should only be mentioned in the article text (e.g., A. Kingori, University of Nairobi, Kenya, personal communication). Journal names are abbreviated according to Chemical Abstracts. Authors are fully responsible for the accuracy of the references.

Examples:

Chikere CB, Omoni VT and Chikere BO (2008). Distribution of potential nosocomial pathogens in a hospital environment. *Afr. J. Biotechnol.* 7: 3535-3539.

Moran GJ, Amii RN, Abrahamian FM, Talan DA (2005). Methicillinresistant *Staphylococcus aureus* in community-acquired skin infections. *Emerg. Infect. Dis.* 11: 928-930.

Pitout JDD, Church DL, Gregson DB, Chow BL, McCracken M, Mulvey M, Laupland KB (2007). Molecular epidemiology of CTXM-producing *Escherichia coli* in the Calgary Health Region: emergence of CTX-M-15-producing isolates. *Antimicrob. Agents Chemother.* 51: 1281-1286.

Pelczar JR, Harley JP, Klein DA (1993). *Microbiology: Concepts and Applications.* McGraw-Hill Inc., New York, pp. 591-603.

Short Communications

Short Communications are limited to a maximum of two figures and one table. They should present a complete study that is more limited in scope than is found in full-length papers. The items of manuscript preparation listed above apply to Short Communications with the following differences: (1) Abstracts are limited to 100 words; (2) instead of a separate Materials and Methods section, experimental procedures may be incorporated into Figure Legends and Table footnotes; (3) Results and Discussion should be combined into a single section.

Proofs and Reprints: Electronic proofs will be sent (e-mail attachment) to the corresponding author as a PDF file. Page proofs are considered to be the final version of the manuscript. With the exception of typographical or minor clerical errors, no changes will be made in the manuscript at the proof stage.

Fees and Charges: Authors are required to pay a \$550 handling fee. Publication of an article in the Journal of Parasitology and Vector Biology is not contingent upon the author's ability to pay the charges. Neither is acceptance to pay the handling fee a guarantee that the paper will be accepted for publication. Authors may still request (in advance) that the editorial office waive some of the handling fee under special circumstances.

Copyright: © 2012, Academic Journals.

All rights Reserved. In accessing this journal, you agree that you will access the contents for your own personal use but not for any commercial use. Any use and or copies of this Journal in whole or in part must include the customary bibliographic citation, including author attribution, date and article title.

Submission of a manuscript implies: that the work described has not been published before (except in the form of an abstract or as part of a published lecture, or thesis) that it is not under consideration for publication elsewhere; that if and when the manuscript is accepted for publication, the authors agree to automatic transfer of the copyright to the publisher.

Disclaimer of Warranties

In no event shall Academic Journals be liable for any special, incidental, indirect, or consequential damages of any kind arising out of or in connection with the use of the articles or other material derived from the JPVB, whether or not advised of the possibility of damage, and on any theory of liability.

This publication is provided "as is" without warranty of any kind, either expressed or implied, including, but not limited to, the implied warranties of merchantability, fitness for a particular purpose, or non-infringement. Descriptions of, or references to, products or publications does not imply endorsement of that product or publication. While every effort is made by Academic Journals to see that no inaccurate or misleading data, opinion or statements appear in this publication, they wish to make it clear that the data and opinions appearing in the articles and advertisements herein are the responsibility of the contributor or advertiser concerned. Academic Journals makes no warranty of any kind, either express or implied, regarding the quality, accuracy, availability, or validity of the data or information in this publication or of any other publication to which it may be linked.

Journal of Parasitology and Vector Biology

Table of Content: Volume 5 Number 4 April 2013

ARTICLES

- Reuse of experimental huts for indoor residual spraying is feasible** 36
Seth Irish, Raphael N'Guessan, Abibatou Odjo, Clemence Metonnou,
Pelagie Boko, Martin
- Prevalence of lungworm infection in small ruminants in North Gondar zone
Amhara National Regional State, Ethiopia** 40
Yitagele Terefe, Ketema Tafess, Getasew Fekadie, Nigatu Kebede
- Two avian cestodes parasitic to Corvus species of Kashmir, India** 46
Javid Ahmad Dar, Syed Tanveer

Full Length Research Paper

Reuse of experimental huts for indoor residual spraying is feasible

Seth Irish^{1,3*}, Raphael N'Guessan^{1,2}, Abibatou Odjo^{2,3}, Clemence Metonnou^{2,3},
Pelagie Boko^{2,3}, Martin Akogbeto² and Mark Rowland^{1,2,3}

¹London School of Hygiene and Tropical Medicine, Keppel Street, London WC1E 7HT, UK.

²Centre de Recherche Entomologique de Cotonou, 06 BP 2604, Cotonou, Benin.

³Pan-African Malaria Vector Research Consortium, Moshi, Tanzania and Cotonou, Benin.

Accepted 26 March, 2013

Experimental huts are costly investments for programs evaluating insecticides and researching mosquito behaviour. These huts can be used to evaluate indoor residual sprays for malaria control but when the hut trials finish, contamination by the long lasting insecticide treatments might prevent further use of the hut. To see if experimental huts could be reused after indoor residual spraying tests, huts in Cotonou, Benin, were treated with a high dose of chlorpyrifos methyl (500 mg/m²). Bioassays with susceptible *Anopheles gambiae* indicated the treatment was successful. After this, untreated surfaces were washed and the inner surface of the cement walls was chipped away and replaced. Bioassays indicated that contamination was not present and that reuse of huts after indoor residual spraying is possible.

Key words: *Anopheles gambiae*, experimental huts, reuse, indoor residual spraying, Benin, chlorpyrifos methyl.

INTRODUCTION

Indoor Residual Spraying (IRS) has been an effective tool in the control of malaria vectors. In India, the number of cases of malaria was dramatically decreased, from 75,000,000 cases in 1947 to 100,000 cases in 1965, after 7 years of eradication operations, largely through IRS programs (Sharma, 1987). Significant reductions were also attributed to IRS programs in Kenya and Tanzania (Draper and Smith, 1962). The primary vector control intervention at the moment is insecticide treated nets (ITN), which are being widely distributed, especially for children and pregnant mothers. However, indoor residual spraying is also being rapidly scaled up; the number of people protected by IRS increased from 15 million in 2006 to 59 million in 2008 (WHO, 2009).

The World Health Organization (WHO) has provided guidelines for the testing of indoor residual sprays (WHO, 2006). These include three test phases. Phase I is composed of bioassays in the laboratory. Phase II trials are carried out in experimental huts, and Phase III evaluations are large-scale field trials. In addition to IRS treatments, evaluations of insecticides, repellents, and even behavioural studies can be tested in experimental huts (WHO, 2006; Suwonkerd et al., 2006).

Experimental huts are small houses with entrances allowing mosquitoes to enter but not easily leave. Thus all mosquitoes entering the hut can be counted and scored the following morning as alive or dead, and bloodfed or un-fed. The addition of a veranda allows induced

*Corresponding author. E-mail: seth.irish@lshtm.ac.uk. Fax: +44 (0)20 7436 5389.

exophily to be measured as mosquitoes try to distance themselves from a treatment. Hougard et al. (2007) described the type of experimental hut used in this study.

As experimental huts are expensive to build and maintain, it is important to know whether or not the huts can be reused after treating the walls with a residual insecticide. Any contamination of the walls or other structures could result in misleading results in later tests. The WHO recommends a complete refurbishment of huts between trials to prevent this contamination (WHO, 2006). The feasibility of reusing huts by removing and replacing the inner surface of the walls was tested in Cotonou, Benin. Mortality of *An. gambiae* in cone bioassays was used as a measure of contamination.

MATERIALS AND METHODS

Three experimental huts were selected for use in Ladji, a neighbourhood in Cotonou, Benin (6°23'23N, 2°25'56E). Ladji is on the edge of Lake Nokoué and has experimental huts that have been in use, primarily for testing insecticide treated mosquito nets, since 2002 (N'Guessan et al., 2010).

An initial series of bioassays was conducted to determine the mortality of mosquitoes exposed to various surfaces in the huts before treatment. Bioassays were performed in WHO cones attached to the surfaces with masking tape. Surfaces tested included doors (2 cones), walls (10 cones), entry window slits (2 cones), ceiling (2 cones), floors (2 cones), and verandas (2 cones). The locations of the cones were in random locations for the treated walls and very near the areas treated for untreated surfaces. Mosquitoes used were female *An. gambiae* s.s. Kisumu (a pyrethroid-susceptible laboratory strain originally from Kenya) between two and three days old. Five mosquitoes were put into each cone for 30 min. After this period, they were removed from the cone and put into plastic cups covered with untreated mosquito netting and given access to 10% honey solution. Mortality was scored after 24 h.

After the initial survey, two huts were treated with the organophosphate chlorpyrifos methyl ('Reldan GF 1246', Dow AgroSciences) at a dose of 500 mg/m². This was intended to represent a highly effective dose for indoor residual treatments for this insecticide. Ansari and Razdan (2004) came to the conclusion that an IRS treatment of 500 mg/m² gave high efficacy in controlling *Anopheles culicifacies*. N'Guessan et al. (2010) found chlorpyrifos methyl (500 mg/m²) to provide high levels of control for a longer period than DDT (2 g/m²) and the pyrethroid, lambda-cyhalothrin (30 mg/m²). It is estimated that this dose would give a good idea if any of the insecticide was remaining after the resurfacing of the cement walls. The window slits that allow mosquitoes to enter, the metal door, the plastic ceiling, the floor near the wall, and the veranda were not treated and these surfaces were covered with plastic sheeting in one treatment hut (Hut 1), and with cement bag paper in the other (Hut 2). Reed mats were treated and nailed to the ceiling. Treatment was performed using a Vexmorel 2000 Pro (Berthoud, Villefranche, France) backpack sprayer. Five days after the treatment, bioassays were repeated following the same procedures.

After these bioassays, the interior structures of the huts, including the floors, were washed using laundry detergent (Omo; Unilever Nigeria, Abuja, Nigeria). The surface of inner walls of the treated

rooms (1 to 2 cm, not including the veranda) was chipped away by a mason. They were then resurfaced using a cement and sand mixture.

The walls were left to dry for four days. After this period, the original bioassays were repeated using *An. gambiae* (Kisumu) of the same age (2 to 3 days).

The numbers of dead mosquitoes compared to total numbers were analysed using blocked logistic regression, using Stata 8.1 (Stata Co., College Station, Texas, USA).

RESULTS AND DISCUSSION

The mortality of mosquitoes on the different interior surfaces of the huts is shown in Table 1. Before the treatment, mortality was low in all huts, as expected. Huts 1 and 2 did not have any significant differences in overall mortality relative to the control hut ($p > 0.05$).

All mosquitoes tested in the huts treated with chlorpyrifos methyl at 500 mg/m² died, whether they were exposed to treated surfaces or not. This was not expected as many of the surfaces tested had been covered during the treatment. It was unlikely that this contamination of untreated structures came from a faulty covering with the plastic sheeting or cement bags, as these were well secured. Mosquitoes that were held in cups in the room during testing but were not exposed to treated surfaces were also found dead after 24 h. This indicates there was probably another cause of the mortality of the mosquitoes than a direct contamination of these structures. Chlorpyrifos methyl has a low vapour pressure which may have contributed to the mortality of the mosquitoes, particularly as the tests were done only five days after the initial treatment. The huts were also closed after the treatments and not opened until the time of testing. A vapour effect in the veranda is worrying as it may result in an increased number of dead mosquitoes the verandas. Bar-Zeev and Self (1966) found greater mortality of mosquitoes in window traps on huts treated with propoxur and bromphos than in window traps on control huts, though in both cases the mosquitoes were not in contact with the treated surfaces indoors. As the main experimental hut models all use window traps or verandas to monitor mortality in exiting mosquitoes, this could lead to an overestimation of the insecticidal effect on mosquitoes that come into contact with treated walls and attempt to leave the hut.

There was little (less than 10%) mortality in mosquitoes tested on surfaces in the control hut was at the same time as the treated huts (shown in Table 1 as "after treatment").

After the removal of the inner surface of the walls and its replacement, the surfaces were tested again. There was some mortality (<15%) on the walls of Hut 2, and in the control, so these tests were repeated to see if this was mortality due to the treatment or not. In the end, no

Table 1. Results of WHO cone tests using *Anopheles gambiae* (Kisumu) in experimental huts in Ladji, Cotonou (Benin).

Test condition	Percentage mortality (number tested)					
	Walls	Doors	Window slits	Floor	Ceiling	Veranda
Hut 1 (plastic covering)						
Before treatment	2.0 (49)	0 (10)	0 (10)	10 (10)	-	0 (9)
After treatment	100 (50)	100 (10)	100 (10)	100 (10)	100 (9)	100 (10)
After refection of walls	1.0 (105)	0 (23)	0 (20)	5.0 (20)	0 (9)	4.2 (24)
Hut 2 (cement bag covering)						
Before treatment	0 (53)	0 (9)	0 (8)	12.5 (8)	0 (4)	11.1 (9)
After treatment	100 (47)	100 (10)	100 (8)	100 (10)	100 (9)	100 (10)
After refection of walls	3.7 (215)	5 (20)	4.2 (24)	4.2 (24)	0 (11)	4.3 (23)
Control						
Before treatment	0 (50)	0 (8)	0 (10)	0 (7)	0 (1)	0 (9)
After treatment	0 (46)	0 (12)	0 (11)	9 (11)	0 (10)	0 (10)
After refection of walls	1.9 (105)	9.1 (33)	4.5 (22)	0 (24)	4.8 (21)	0 (22)

hut had mortality greater than 10% on any surface. The results of mortality on covered structures for the hut with plastic coverings and the hut with cement bag coverings were not different before and after treatment ($p=0.255$). The plastic covering seemed to allow a better covering of surfaces to be left untreated. However, the run-off of the spray that happened to touch the plastic ran down the plastic sheeting more easily than on the cement bags, which absorbed some of the spray. It is important to keep this in mind while spraying to avoid having these drops touch surfaces that are not to be treated.

Before removing the inner layer of cement from the walls, the huts were vigorously cleaned. The plastic ceilings were not protected during spraying but were cleaned with soap and water. The fact that the ceilings caused no mortality in the final tests shows the impact of cleaning with soap and water.

Conclusions

The results from the treated huts indicate that reuse of experimental huts is possible, as the results of all the tests before the treatment and after the refection were not significantly different. The most important steps in reuse of experimental huts are: good covering of structures to be protected, vigorous cleaning of these structures before the refection of the walls (to avoid contaminating the new walls), a complete removal of the first layer of cement, and a proper refection of walls. The findings from this study provide conclusive evidence that proper refection of huts allows for their reuse, even with highly insecticidal

and long lasting products.

ACKNOWLEDGEMENTS

This work was funded by the Innovative Vector Control Consortium. Thanks to Janet Hemmingway for suggesting this exercise. The anonymous reviewers are also thanked for their comments.

REFERENCES

- Ansari MA, Razdan RK (2004). Impact of residual spraying of Reldan against *Anopheles culicifacies* in selected villages of District Ghaziabad (Uttar Pradesh), India. J. Vector Borne Dis. 3-4:54-60.
- Bar-Zeev M, Self LF (1966). A note on the use of window-traps as a tool for evaluating insecticides. Mosq. News 26:205-207.
- Draper CC, Smith A (1962). Malaria in the Pare area of Tanganyika. Part II. Effects of three years' spraying of huts with dieldrin. Roy. Soc. Trop. Med. Hyg. 54:342-357.
- Hougard JM, Martin T, Guillet PF, Coosemans M, Itoh T, Akogbeto M, Chandre F (2007). Preliminary field testing of a long-lasting insecticide-treated hammock against *Anopheles gambiae* and *Mansonia* spp. (Diptera: Culicidae) in West Africa. J. Med. Entomol. 44:651-655.
- N'Guessan R, Boko P, Odjo A, Chabi J, Akogbeto M, Rowland M (2010). Control of pyrethroid and DDT-resistant *Anopheles gambiae* by application of indoor residual spraying or mosquito nets treated with a long-lasting organophosphate insecticide, chlorpyrifos-methyl. Malar. J. 9: 44.
- Sharma, GK (1987). A critical review of the impact of insecticidal spraying under NMEP on the malaria situation in India. J. Commun. Dis.19: 187-290.
- Suwonkerd W, Mongkalagoon P, Parbaripai A, Grieco J, Achee N, Roberts D, Chareonviriyaphap T. (2006). The effect of host type on movement patterns of *Aedes aegypti* (Diptera: Culicidae) into and out

of experimental huts in Thailand. *J. Vector Ecol.* 31: 311-318.
World Health Organization (2006). Guidelines for Testing Mosquito
Adulticides for Indoor Residual Spraying and Treatment of Mosquito
Nets. WHO/CDS/NTD/WHOPES/GCDPP/2006.3

World Health Organization (2009). World Malaria Report 2009. Geneva,
Switzerland.

Full Length Research Paper

Prevalence of lungworm infection in small ruminants in North Gondar zone, Amhara National Regional State, Ethiopia

Yitagele Terefe¹, Ketema Tafess², Getasew Fekadie¹ and Nigatu Kebede^{3*}

¹Faculty of Veterinary Medicine, Haramaya University, Ethiopia.

²Faculty of Veterinary Medicine, Gondar University, Gondar, Ethiopia.

³Aklilu Lemma Institute of Pathobiology, Addis Ababa University, Addis Ababa, Ethiopia.

Accepted 15 March, 2013

The study was conducted to determine the prevalence, associated risk factors and identification of the species of lungworms of small ruminants in four districts of Northwestern Amhara National Regional State. A total of 632 small ruminants were included in the study using coprologic procedures (565 animals) and post-mortem examination (87 animals). The study showed that the overall prevalence of lungworm infection was 46.0 and 56.3% based on coprological and postmortem examination, respectively. Significant difference ($P < 0.05$) in prevalence of lungworm was found between animals under semi-intensive and extensive management systems. The prevalence of lungworm infection was significantly different ($P < 0.05$) between seasons, the highest being during the rainy season (57.1%) and the lowest in the dry season months (35.0%). The prevalence of *Muelleris capillaris* infection was highest (50.4%) during the study period based on postmortem examination. The prevalence of lungworm infection showed significant association ($P < 0.05$) with different age groups of animals, where *Dictiocaulus fillaria* was highly prevalent in young age group while *M. capillaris* and *Protostrongylus rufescens* were highly prevalent in adult age groups of study animals. In the current study, three respiratory helminthes of small ruminants were identified and management system, age and season are important risk factors associated with lungworm infection in the study area. Due to its impact on production, emphasis should be given for the prevention and control of lungworm infection in the study area.

Key words: *Dictiocaulus fillaria*, *Muelleris capillaries*, *Protostrongylus rufescens*, lungworm, small ruminant, helminthes.

INTRODUCTION

Sheep and goats are the most numerous of man's domestic livestock and are especially important in the more extreme climates. Of the worlds' 1,614 million sheep and 475 million goats, 65 and 95%, respectively, are located in developing countries. Small ruminants in Africa are noted for their ability to convert low cost feed into high

value products, namely: meat, milk, fiber, manure and hides (Wilsmore, 2006; Food and Agriculture Organization (FAO), 1986).

The current sheep and goat populations of Ethiopia are approximately 23 and 17 million, respectively (ILRL, 2000). Small ruminants are important contributors to food

*Corresponding author. E-mail: nigatukebede@yahoo.com. Tel: +251112763091. Fax: 251-11-2755296.

production in Ethiopia, providing 33% of meat consumption and 14% of milk consumption (Asfaw, 1997). In the central highlands of Ethiopia where mixed crop-livestock production system is practiced, small ruminants account for 40% of cash income and 19% of the house hold meat consumption (Zelalem and Fletcher, 1993). Owing to their high fertility, short generation interval and adaptation even in harsh environments, sheep and goats are considered as investments and insurance to provide income to purchase food during seasons of crop failure and to meet seasonal purchases such as improved seed, fertilizer and medicine for rural households (Asfaw, 1997).

Even though small ruminants are important components of the Ethiopian farming system, their contribution to food production, rural income and export revenue are far below than their expected potential. This is because small ruminant production is constrained by the compound effects of disease, poor feeding and poor management (Getachew, 1995). Studies in the central highland of Ethiopia have shown that lungworm parasites are a major problem in small ruminants and cause disease, increase mortality and production losses (Bekele et al., 1992). Up to half of all sheep deaths and morbidity on farms in Ethiopian highlands are caused by parasitic pneumonia and endoparasites (ILCA, 1990).

The incidence of lungworm infection varies geographically. Only a few limited studies have been completed on respiratory helminthes of small ruminants in the Amhara National Regional State, northwestern Ethiopia. Therefore, the aims of the present study are (i) to determine the prevalence of lungworm infection in small ruminants; (ii) to identify the species of lungworms and (iii) to identify associated risk factors for lungworm infection in north Gondar zone, Amhara National Regional State, northwestern Ethiopia.

MATERIALS AND METHODS

Study area

The study was conducted in four districts of northwest Amhara region including Abaintonios, Anchew, Belagig and Gondar town districts. The study areas are about 710 km North West of Addis Ababa at an altitude ranging from 1500 to 3500 meters above sea level. Numerous mountains, plateaus, sloped areas, rivers, streams and lakes mark the topography of the area. The climate of the study area is divided into subhumid ("wurch and Dega"), semiarid ("Woynadega") and arid ("Kola"). The average maximum and minimum daily temperature of the area varies between 22 to 30.7°C and 12.3°C, respectively.

The region receives a bimodal rainfall, the average annual precipitation rate being 1000 mm. The short rains occur during the months of March to May while the long rains extend from June to September. The production system observed in the area combines cereal-based agriculture and livestock farming. The study area has a livestock population of 2.03 million cattle, 0.6 million sheep, 0.54 million goats, 0.25 million equine species of which 62.5% are donkeys, 5% are mules, and 32.5% are horses, and 1.9 million poultry (CSA, 2003).

Study animals and sampling

Small ruminants in the study area are kept mainly under an extensive traditional management system, involving small household flocks of mixed age. The sample size was determined according to Thrusfield (1995) for an infinite population. Since there has been no previous work done in the study area, an estimated prevalence of 50% was used for sample size determination. A total of 652 small ruminants (412 sheep and 153 goats from study districts, and 58 sheep and 29 goats from Gondar ELFORA abattoir) were used in this study. Four representative districts were purposely selected; Abaintoniose, Anchew, Blagig and Gondar town. Sampling animals was conducted at random from study districts and small ruminants slaughtered in ELFORA abattoir. Animals from Gondar town were managed under a semi-intensive system while the rest were under an extensive production system.

Study design

Coprological examination

Faecal samples were collected directly from the rectum of all selected animals and stored in vials containing 10% formalin and transported to the Gondar veterinary clinic for laboratory examination. In the laboratory, the collected fecal samples were processed according to conventional methods for lungworm larvae (Baermanization) (Hansen and Perry, 1994). During sample collection, the species of the animal, sex, age, date of sampling and the area were recorded. In the laboratory, 25 g of faeces was weighed from each sample for the extraction of larvae using modified Baermann technique (Hansen and Perry, 1994). The faeces were enclosed in a gauze fixed on the string rod and submerged in a clean glass tube filled with warm water. The whole apparatus was left for 2 to 3 h. The larva leaves the faeces and migrates through the gauze and settles at the bottom of the glass. After siphoning off the supernatant, the sediment was examined under the low power of the microscope (Fraser, 1991; Urquhart et al., 1996). All the area under the cover slip was thoroughly and uniformly searched for the presence of lungworm larvae (Hendrix, 1998).

Abattoir survey

Small ruminants slaughtered at Gondar ELFORA abattoir were examined during the study period. The lungs were palpated for the presence of protostrongylidea nodules. These are brood nodules and worm nodules. Brood nodules are cone shaped granuloma-like areas of the affected lung tissue varying in size and color, which contains active worm and caused by parenchyma-dwelling small lungworms (Protostrongylidae). The worm nodules are pin head shaped mostly subpleural, grey (*Mullerius*) cyst which contain adult worms. If the nodules are present, they are trimmed off and worms extracted from the tissue by gentle pressing of a small non-calcified nodule or part of large nodule between two glass slides and then carefully teasing the worm away from the tissue. The air passages were opened starting from the trachea down to the small bronchi with fine blunt pointed scissors, to detect parasites (Kassai, 1999; Schneider, 2000).

Data analysis

Prevalence was determined by the percentage (%) positive and chi-square (χ^2). To measure association were

Table 1. Prevalence of lungworm infection in different districts of the study area.

District	Number examined	Number positive (%)
Abaintonios	176	77 (43.7)
Anchew	88	50 (56.8)
Belagig	56	24 (42.8)
Gondar town	245	109 (44.4)
Total	565	260 (46.0)

P > 0.05.

the statistical tools applied and determined using statistical package for social sciences (SPSS) V 17.0. In all the analysis, confidence level was held at 95% and 5% level of significance (Thursfield, 1995).

RESULT

Coprological examination

Out of 565 (412 sheep and 153 goats) fecal sample examined, 260 (46.0%) were positive for lungworm infection. No statistically significant difference was observed in the prevalence of the lungworm infection of small ruminants in different district ($\chi^2 = 28.22$ P > 0.05). The higher prevalence rate was encountered in Anchew (56.8%) while the lowest was in Belagig (42.8%) (Table 1).

The prevalence rate of lungworm infection was found to be higher in extensive management system (51.8%) than in semi-intensive management system (24.5%). Comparison between sheep and goats showed statically significant difference between the two species ($\chi^2 = 33.2$, P < 0.05). Similarly, there was association between the prevalence rate of lungworm infection with management system, and months where $\chi^2 = 36.71$, P < 0.0 and $\chi^2 = 47.55$ P < 0.05, respectively. The prevalence rate of lungworm infection did not show statistically significant association with sex of the animals ($\chi^2 = 4.55$, P > 0.05). Lungworm infection in small ruminants and associated risk factors in the study area are shown in Table 2.

In comparison of age groups, significant difference ($\chi^2 = 33.22$, P < 0.05) in the prevalence rate of lungworm infection was observed. Among lungworms identified, high prevalence rate of *Muelleris capillaries* (50.4%) followed by *Dictyocaulus fillaria* (26.5%) was recorded. The prevalence of *M. capillaris* and *Protostrongylus rufescens* increases with age of the animal, whereas the prevalence of *D. filaria* was high in young age groups of the animals. Mixed infection was common in old age group of the animals (Table 3).

Abattoir survey

Of the 87 animals (58 sheep and 29 goats) examined in the ELFORA abattoir, 49 (56.3%) were positive for

lungworm infection. There was a significant difference in prevalence of lung worm infection between the two species ($\chi^2 = 3.95$, P < 0.05), the prevalence being higher in sheep (63.8%) than goats (41.3%). The distribution of lungworm species detected in the lung during postmortem examination of positive animals is shown in Table 4. Higher prevalence of *M. capillaris* (51.0%) was observed than *P. rufescens* (28.6%) and *D. fillaria* (20.4%). The relative percentage of adult lungworms recovered during postmortem examination is described in Table 4.

DISCUSSION

This study revealed the overall prevalence of lungworm infection of small ruminants to be 46.0%, with 42.3% in sheep and 46.7% in goats. It also disclosed that animals are infected with three nematode species parasitizing the respiratory tract of small ruminants of which *Mullerius* and *Dictyocaulus* species are the most abundant. This finding is in agreement with Regassa et al. (2010) and Sissay (1996) who have reported 40.4 and 44% in small ruminants in Dessie and Kombolcha districts, north-eastern Ethiopia and Bahir Dar, respectively. The present finding is lower than reports of Alemu et al. (2006) in northeast Ethiopia and Bekele and Abu (2011) in Tiyo District, South-East Ethiopia, with an observed prevalence of 53.6 and 57.1%, respectively. However, it is higher than that of Addis et al. (2011) and Weldesenebet and Mohamed (2012), with 33.83% prevalence at Gondar and 26.7% at Jimma, respectively. These differences might be due to the method used for the detection of the larvae or difference in the study area of topography which has a conducive environment for the survival of larvae and intermediate hosts, slug or snails and also the nutritional detection.

In this study, small ruminants under extensive management system had more infection than those kept under semi-intensive management system. The reason for this could be increased cultivation of land which restricts animals on communal grazing land so that large numbers of the animals are kept together. This could increase the degree of pasture contamination leading to higher prevalence rate. Management practice such as provision of ample nutrition increases the resistance of the host under the semi-intensive system, contrary to this malnutrition which reduces the host-parasite response and favors the fecundity of the parasites that allows the animals for continuous larvae exposure under extensive system (Soulsby, 1986).

The reason for this high prevalence of *Mullerius* compared with *Dictyocaulus* could be partly attributable to its wide range of intermediate host and the ability of larvae to over winter in the mulluscs. Additional factors which play a part in ensuring the endemicity of the worm are, first, the ability of L1 (First stage larva) to survive for months in faecal pellets and secondly, the persistence of

Table 2. Lungworm infection in small ruminants and associated risk factors in the study area.

Variable	Number examined	Number positive (%)
Species		
Ovine	412	164(39.8)
Caprine	153	96(62.7)
Sex		
Female	329	158(48.0)
Male	236	102(43.2)
Management system		
Extensive	444	230(51.8)
Semi-intensive	121	30(24.5)
Season		
Dry	285	100(35.0)
Rainy	280	160(57.1)

Table 3. Prevalence of the species of lungworm in small ruminants of different age groups.

Age (years)	Prevalence (%)	Positive							Total
		Single infection			Mixed infection				
		D (%)	M (%)	P (%)	DM (%)	DP (%)	MP (%)	DMP (%)	
≤1/2	29(11.6)	14(20.3)	12(9.2)	2(4.9)	-	-	1(14.3)	-	81
1/2-2	80(30.8)	24(34.9)	41(31.3)	10(24.4)	2(33.3)	1(25.0)	2(28.6)	-	193
2-4	69.(26.5)	12(17.4)	41(31.3)	11(26.8)	3(50.0)	1(25.0)	1(14.3)	-	146
>4	82(31.5)	19(27.4)	37(28.2)	18(43.9)	1(16.7)	2(50.0)	3(42.9)	2(100.0)	145
Total	260(46.01)	69(26.5)	69(26.5)	41(15.8)	6(2.3)	4(1.5)	7(2.7)	2(0.8)	565

D: *Dictyocaulus filaria*, M: *Muellerius capillaris*, P: *Protostrongylus rufescens*; $\chi^2 = 33.72$, $P < 0.05$.

Table 4. Relative percentage of adult lungworm recovered from postmortem examination.

Species of lung worm	Number of positive	Percentage (%)
<i>D. filarial</i>	10	20.4
<i>M. capilaris</i>	25	51.0
<i>P. rufescens</i>	14	28.6
Total	49	100.0

$\chi^2 = 87.0$, $P < 0.05$.

the L3 (third stage larva) in the intermediate host for the life time of the mollusks (Taylor et al., 2007). On the other hand, the longevity and development of free larvae of *Dictyocaulus* are known to be dependent on humidity and temperature condition. Dry seasons are characterized by high mortality of larvae in the pasture (Gallia and Nunns, 1976), where dry, hot summer and cold winter is the climatic condition of the study area. The prevalence of *D. filaria* is higher in infants than adults. However, the prevalence of *M. capillaris* and *P. rufescens* is higher in adult animals than younger ones. This might be due to the long

period of potency and the apparent inability of the final host to develop acquired immunity, so that adult sheep have the heaviest infection, highest infection and the highest prevalence (Taylor et al., 2007). The study further revealed that goats were found to be more susceptible to lungworm infection than sheep. This could be because most of the goats in this study were from lowland and mid altitude areas, which are thought to be suitable for survival of the larval stage of the parasite. Likewise, in the lowland areas of the country where goats are mostly reared, there is poor veterinary infrastructure and medication.

More importantly, the condition could be due to less or slow development of immunity in goats compared to sheep (Wilsmore, 2006).

The higher prevalence of lungworm infection during the rainy season of the months might be the presence of sufficient moisture during the rainy season which favored the survival of infective larvae and stimulated the activity of mollusks in the pasture resulting in higher probability of up take of the infective larvae, leading to higher prevalence rate. In northwestern part of Ethiopia, under local production system, the animals that are completely managed on pasture grazing throughout the year succumb to seasonal variation of availability of forgeable feed and then difference in plane of nutrition. Thus, the presence of sufficient feed in rainy season could in turn increase the nutritional status, and these well fed animals develop good immunity that suppressed the fecundity of the parasites. This situation was reported by Bisset et al. (1996) that increased plane of nutrition increased the immunity and reduced the fecundity of the worm, contrary to the higher prevalence rate observed during the rainy season.

The prevalence of lungworm infection obtained by post-mortem examination was higher (56.3%) than the result obtained by coproscopic examination (46.0%). This difference might be related to worm nodes of Protostrongylidae. Worm nodules contain immature parasite in general (Schneider, 2000). Higher infection rate observed on postmortem examination may be related to these worm nodules detected. In addition, in the prepatent or postpatent phase or during hypobiosis, it is impossible to detect this parasite by fecal examination (Fraser, 1991). Furthermore, egg laying adult female parasites might be inhibited by the immune reaction of the host (Hansen and Perry, 1994).

Conclusion

The study revealed that lungworm infection in small ruminant is highly prevalent in North Gondar zone. *D. filaria*, *M. capillaris* and *P. rufescens* were the lungworms identified during the study. Young animals are mostly affected by most pathogenic species of lungworm, *D. filarial*, than adults on the contrary which are highly affected by *M. capillaris* and *P. rufacens*. Animals under semi-intensive management system have been found less affected than those under extensive management system. Therefore, avoidance of overstocking and grazing in watery and damp areas of arid pastures, treatment of potential worm carriers and separate grazing of young stocks are recommended.

REFERENCES

Addis M, Fromsa A, Ebuy Y (2011). Study on the Prevalence of Lung worm Infection in Small ruminants in Gondar Town, Ethiopia. Vet. Res. 4(3):85-89.

Alemu S, Leykun EG, Ayelet G, Zeleke A (2006). Study on small

ruminants lung worms in northeastern Ethiopia. Vet. Parasitol. 142(2-3):330-335.

Asfaw W (1997). Country report, Ethiopia. In: proceeding of a seminar on livestock Development policies in Eastern and Southern Africa, 28th July-1st August, 1997, Mbabany. Organized by CTA, OAU/IBAR, the Ministry of Agriculture Cooperative, Swaziland.

Bekele M, Abu A (2011). Ovine lungworms in Tiyo District, South-East Ethiopia: Prevalence, effect of altitude and major host related risk factors. Glob. Vet. 7(3):219-225.

Bekele T, Woldeab T, Lahlou-lasso A, Sherington J (1992). Factors affecting Morbidity on-farm and on-station in Ethiopian highland sheep. Acta. Trop. 52(2-3):99-109.

Bisset ST, Thomsborg SM, Mainqi N, Munyua WK (1996). Nematode burdens and immunological responses following natural challenges in Romney lambs selectively breed for low or high fecal worm egg count. Vet. Parasitol. 61(3-4):249-263.

Central Stastical Authority (CSA) (2003). Ethiopian agricultural sample Enumeration report on livestock and farm implement part IV. Addis Ababa, Ethiopia. pp. 29-136.

FAO (1986). Small ruminant production in the developing countries. In: proceedings of an Expert consultation held in Sofia, Bulgaria, 8-12 July 1985 (<http://www.fao.org/docrep/009/ah221e/ah221e00.htm>).

Fraser CM (1991). The Merk veterinary manual. A hand book of diagnosis, therapy and disease prevention and control for the veterinarians, 7th ed. Mark and Co. Inc., Rahway, Nit, USA. pp. 714-614.

Gallie GJ, Nunns VJ (1976). The bionomics of free living larvae and transmission of *Dictyocaulus filarial* between lambs in North-East England. J. Helminthol. 50(2):79-89.

Getachew MR (1995). Parasite of small ruminants. In: Gray GD, Uilengerg G (eds), Parasitological Research in Africa

Hansen J, Perry B (1994). The Epidemiology, Diagnosis and Control of Helminth parasites of ruminants. ILRAD, Nairobi, Kenya. P 38.

Hendrix C (1998). Diagnostic of veterinary parasitology, 2nd ed. Mosby, London. p 49.

International Livestock Center for Africa (ILCA) (1990). Annual report, 1989. (International Livestock Center for Africa, Addis Ababa, Ethiopia. p 37.

International Livestock Research Institute (ILRI) (2000). Hand book of livestock statistics for developing counties. Socio-economics and policy research working paper No. 26. ILRI Nairobi, Kenya. p 299.

Kassai T (1999). Veterinary Helminthological. Butterworth helminthology, Budapest, Hungary. pp. 85-93.

Lebbie SHB, Rey B, Irungu EK (1994). Small ruminant research and development in Africa. Proceeding of the second Biennial conference of the African Small Ruminant Research Network. ILCA, Ethiopia. pp. 1-5.

Regassa A, Toyeb M, Abebe R, Megersa B, Mekibib B, Mekuria S, Debela E, Abunna F (2010). Lungworm infection in small ruminants: prevalence and associated risk factors in Dessie and Kombolcha districts, Northeastern Ethiopia. Vet. Parasitol. 169(1-2):144-148.

Schneider T (2000). Helminths of respiratory system. In: Veterinary Medicine Parasitology, Vol. 5. Voll standing auflage.parey Buch vet log. Berlin. pp. 193-198.

Sissay A (1996). Preliminary study on the prevalence of ovine lungworm infection in and around Bahir Dar. DVM thesis, Faculty of Veterinary Medicine, Addis Ababa University, Debre Zeit, Ethiopia. pp. 14-16.

Solusby EJJ (1986). Helminthes, arthropods and protozoa of domesticated animals, 7th ed. Bailliere, Tindall, London. pp. 262-268.

Taylor MA, Coop RL, Wall RL (2007). Parasites of the respiratory system. In: Veterinary parasitology, 3rd ed. Black well publishing Ltd., Oxford, UK. pp. 195-199.

Thrusfield M (1995). Surveys. In: Veterinary Epidemiology, 2nd ed. Black well science Limited, Cambridge, USA. pp. 178-198.

Urquhart HM, Armour Duncan JL, Dunn AM, Jennings FW (1996). Veterinary parasitology, 2nd ed. Black Well science limited, London. pp. 301-309.

Weldesenebet D, Mohamed A (2012). Prevalence of small ruminant lung worm infection in Jimma town. Glob. Vet. 8(2):153-159.

Wilsmore T (2006). Disease of small ruminants in Ethiopia, the veterinary epidemiology and economics research unit of agriculture's policy and development the university of read, UK, pp: 602.

Zelalem A, Fletcher IC (1993). Small ruminant productivity in the central Ethiopia Mixed farming system. In: Proceeding of the 4th National Livestock Improvement Conference, 13-15 November, IAR, Addis Ababa, Ethiopia.

Full Length Research Paper

Two avian cestodes parasitic to *Corvus* species of Kashmir, India

Javid Ahmad Dar* and Syed Tanveer

Department of Zoology, University of Kashmir, Srinagar-190 006, India.

Accepted 18 March, 2013

Three species of birds belonging to the genus *Corvus* (*Corvus monedula*, *Corvus splendens* and *Corvus macrorhynchos*) were collected from nine different localities of Kashmir valley and investigated for the presence of helminthes. *Dilepis undula* (Schrank, 1788) was recovered from all the three host species. While, *Mayhewia kavini* Chishti and Khan, 1982 was recovered only from *C. monedula* and no specimen of this cestode was obtained from *C. splendens* and *C. macrorhynchos* during the present study. The specimens were identified and redescribed on the basis of various morphological and morphometric characters when compared to the known species of genera as *Dilepis* and *Mayhewia*, respectively. However, some intraspecific variations were observed when the present measurements were compared with those of previous authors. The prevalence, mean intensity and abundance of the parasites were determined.

Key words: Cestode, crows, *Corvus*, *Dilepis*, *Mayhewia*, prevalence, Kashmir.

INTRODUCTION

The present study was a part of helminthological investigation carried out on three *Corvus* species (*Corvus monedula*, *Corvus splendens* and *Corvus macrorhynchos*) of Kashmir from November, 2007 to May, 2009. *C. monedula* Linnaeus, 1758 (Jackdaw) is a black-plumaged passerine bird with distinctive white irises; sexes and ages are alike (Goodwin, 1983). It is omnivorous and feeds on plant material and invertebrates (Lockie, 1956). *C. splendens* Vieillot, 1817 (House crow) is about 40 cm in length with lighter grey-brown neck and breast. The wings, tail and legs are black. It appears to be associated with humans and no populations are known to exist independently of humans (Nyari et al., 2006). *C. macrorhynchos* Wagler, 1827 (Jungle crow) is a Large-billed crow with black glossy wings, tail, face and cause diseases include viruses, bacteria, protozoans and throat. The groups of parasites which infect birds and helminthes besides some arthropod ectoparasites. Sexually mature tapeworms live in the intestine or the

diverticula of all classes of vertebrates (Roberts and Janovy, 2005). These parasites are found more frequently in warmer seasons, when intermediate hosts are abundant. Birds become infected by eating an intermediate host (infested with larval stage of a cestode), which may be an insect, crustacean, earthworm, slug, snail or leech depending upon the species of tapeworm (Calnek, 1997). Over 4,000 species of cestodes have been described from animals (Schmidt, 1986). Recently many workers have contributed to the knowledge of avian cestodes such as evaluation of genetic basis of host specificity of cestodes (Benesh, 2010; Henrich et al., 2013), description of ultrastructural characters of the spermiogenesis and mature spermatozoon of *Notopentorchis* sp. by means of transmission electron microscopy (Yoneva et al., 2012), description of some new cestode species like *Spiniglans sharpiloi* from *Pica pica* (Kornyushin et al. 2009), *Cotugnia orientalis* from *Gallus gallus domesticus* (Nanware et al., 2011).

*Corresponding author. E-mail: javid60@gmail.com. Tel: +918803314150.

Table 1. Prevalence, mean intensity and abundance of *Dilepis undula* in *Corvus* species of Kashmir.

Host spp.	NE	NI	NP	P%	MI	AB
<i>C. monedula</i>	30	6	18	20	3	0.6
<i>C. splendens</i>	23	3	11	13.0 4	3.6 6	0.47
<i>C. macrorhynchos</i>	12	3	11	25	3.6 6	0.92
Total	65	12	40	18.4 6	3.3 3	0.61

NE = number examined; NI = number infected; NP = number of parasites; P = prevalence; MI = mean intensity; Ab = Abundance.

Table 2. Prevalence, mean intensity and abundance of *Mayhewia kavini* in *Corvus* species of Kashmir.

Host spp.	NE	NI	NP	P%	MI	AB
<i>C. monedula</i>	30	4	10	13.3	2.5	0.33
<i>C. splendens</i>	23	0	0	0	0	0
<i>C. macrorhynchos</i>	12	0	0	0	0	0
Total	65	4	10	6.15	2.5	0.15

NE = number examined; NI = number infected; NP = number of parasites; P = prevalence; MI = mean intensity; Ab = abundance.

However, systematic knowledge of cestodes from birds of Kashmir including *Corvus*, is still represented through a few references as is obvious by tracing the historical review of cestodes from aves of Kashmir (Gupta, 1967; Fotedar et al., 1970; Chishti, 1973, 1980a, b, c; 1981; Fotedar and Chishti, 1973, 1974, 1976a, b; 1977; Chishti and Khan, 1978; 1979; 1982; Chishti et al., 1986). *Choanotaenia infundibulum*, *Choanotaenia micracantha*, *Anomotaenia galbulae* and *Mayhewia kavini* are the only cestodes reported from *Corvus* species in Kashmir so far (Fotedar and Chishti, 1974; 1976; Chishti and Khan, 1982; Chishti et al., 1986). Thus a thorough investigation of helminth parasitism of *Corvus* species of Kashmir was imperative. The present paper gives redescription of two cestodes (*Dilepis undula* and *M. kavini*) collected from the intestines of three species of crows (*C. monedula*, *C. splendens* and *C. macrorhynchos*) caught from different localities in Kashmir.

MATERIALS AND METHODS

Study area

Kashmir valley is a temperate, North-west Himalayan region of Jammu and Kashmir states in India. It lies between 33° 20' and 34° 54' N latitudes and 73° 55' and 75° 35' E longitudes, covering an area of about 15,948 km². It is a deep bowl shaped valley bounded by lofty mountains of the Pir Panjal and the great Himalayan ranges. The floristic and faunal diversity of the valley is considerably rich owing to its unique topography, temperate climate and geographical isolation from the surrounding plains (Dar et al.,

2002). The birds were collected from different localities of Kashmir valley like Barzulla, Bugam, Chadoora, Khansahib, Naseembagh, Rajbagh, Rambagh, Wathora, and Yousmarg. These collection sites lie within the radius of five to fifty Kilometers (km) from the center of Srinagar city, the summer capital of Jammu and Kashmir States in India and are easily accessible by road transport.

Collection and processing of cestodes

During the present study, 65 birds belonging to three species of *Corvus* (*C. monedula* Linnaeus, 1758; *C. splendens* Vieillot, 1817 and *C. macrorhynchos* Wagler, 1827) were caught alive with the help of nylon net traps, locally known as "Walwash" using suitable baits. The hosts were slaughtered and dissected for parasitological investigation, and the cestode parasites thus collected were fixed in Carnoy's fixative, stained in acetoalum carmine and transferred to xylene for clearing before mounting them in dextrine plasticised xylene (DPX) (Meyer and Olsen, 1975). The drawings of the specimens were made with the help of prism type camera Lucida. Measurements were taken with objective and stage micrometers and expressed in mm (unless otherwise stated). The specimens were identified on the basis of various taxonomic characters using Yamaguti (1961), Schmidt (1986) and Chishti and Khan (1982). Photomicrography was conducted with the help of Digital Olympus Camera.

RESULTS

Two cestode species (*D. undula* and *Mayhewia kavini*) recovered from the intestines of three species of crows (*C. monedula*, *C. splendens* and *C. macrorhynchos*) are

Table 3. Comparative measurements of *Dilepis undula* (Schrank, 1788) with present form (measurements in microns = μ).

Characteristic	Davies (1935)	Matricker (1958)	Chishti (1974)	Present author
No. of rostellar hooks	45-60	48-62	64-70	46-58
Size of outer rostellar hooks (μ)	84	91-116	72-80	74-85
Size of inner rostellar hooks (μ)	72	70-88	60-64	67-78
No. of testes	28-35	28-36	24-26	24-32

redescribed based on their detailed morphological and morphometric studies, revealing some minor intraspecific variations. The prevalence, mean intensity and abundance of each parasite were recorded (Tables 1 and 2).

Dilepis undula (Schrank, 1788)

Hosts: *C. monedula*, *C. splendens* and *C. macrorhynchos*;

Locality: Naseembagh, Rajbagh, Wathora and Yousmarg;

Location: Intestine.

The following redescription is based on twelve cestode specimens, taking four specimens from each *Corvus* species. The cestodes measure 38 to 43 mm in length and 1.36 mm in maximum breadth. All the proglottids are broader and longer, and the posterior margin is broader than anterior margin. The mature proglottids measure 0.18 to 0.27 mm in length and 1.16 to 1.28 mm in breadth. The gravid proglottids measure 0.16 to 0.23 \times 0.8 to 1.25 mm in length and 1.12 to 1.25 mm in breadth (Table 3 and Figure 1: A to E).

The scolex is globular and measures 0.44 to 0.54 mm in length and 0.60 to 0.64 mm in breadth across the suckers. The four suckers are rounded, muscular and measure 0.16 to 0.20 mm in diameter. The rostellum with its sac extends below the posterior margin of suckers and measures 0.44 to 0.58 mm in length and 0.14 to 0.20 mm in maximum breadth; it bears 46 to 58 hooks arranged in a double crown; each crown alternating with the other. The blade of hooks is smaller than handle, with guard as knob-like structure. The hooks of anterior and posterior crown measure 74 to 85 μ and 67 to 78 μ in length, respectively. The scolex is followed by a short neck that measures 0.24 to 0.33 mm in length and 0.50 to 0.55 mm in breadth. The dorsal longitudinal excretory duct is 14 μ in diameter and the ventral duct is 17 μ in diameter. The testes lie posterior to female reproductive organs, numbering 24 to 32 in each proglottid. They are rounded and measure 48 to 53 μ in diameter. The vas-deferens forms many coils in the anterior region of the proglottid. The vesicula seminalis are absent and the ductus ejaculatorius is straight. The cirrus pouch is cortical, extending up to the longitudinal excretory ducts of poral side; measuring 0.12 to 0.15 mm in length and 0.04 to 0.07 mm in width. The cirrus is slender and small 82 to 88 μ in length, armed with small spines. The genital pores

are unilateral and present just near the anterior margin of proglottids, laterally.

The ovary is bilobed, with the poral lobe being smaller than the aporal one; the two lobes show further lobulations. The two lobes are connected in the middle by a narrow U-shaped isthmus. The ovary extends 0.65 to 0.72 mm across. The vitelline gland is compact, horse-shoe shaped and lies posterior to the ovary. It measures 0.12 to 0.17 \times 0.04 to 0.06 mm. An oval receptaculum seminis, 0.08 to 0.10 \times 0.03 to 0.05 mm in size, lies dorsal and anterior to the ovary. The uterus occupies the whole of the gravid proglottid filled with eggs, which measure 23 to 26 \times 14 to 17 μ .

Mayhewia kavini (Chishti and Khan, 1982)

Hosts: *Corvus monedula*;

Locality: Barzulla, Naseembagh, Rambagh and Wathora;

Location: Intestine.

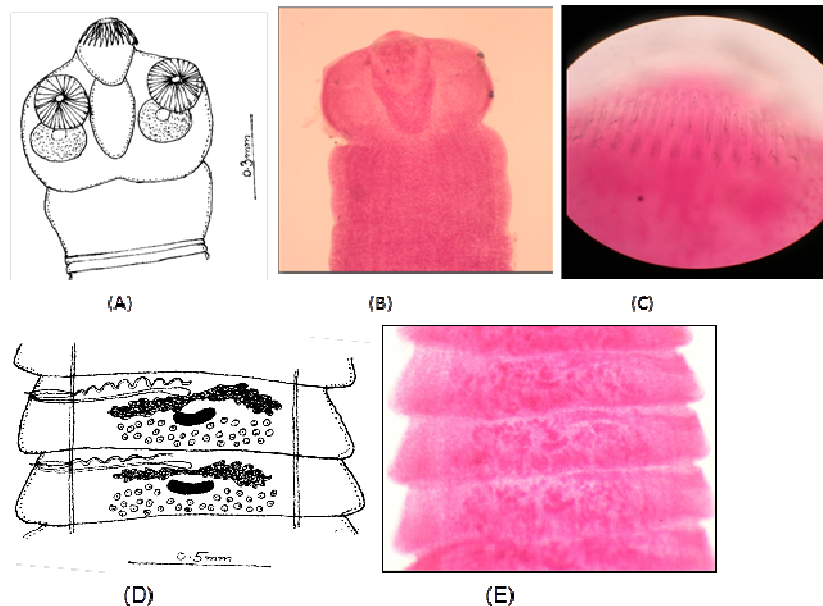
The following redescription is based on ten cestode specimens, collected from the intestines of four, out of thirty jackdaws, *C. monedula*. The cestodes measure 55 to 84 mm in length and 0.77 to 1.0 mm in their maximum breadth. The strobila consists of many proglottids which are broader than being long. The mature proglottids measure 0.20 to 0.26 mm in length and 0.8 to 0.92 mm in breadth. The gravid proglottids are 0.30 to 0.36 mm in length and 0.77 to 1.0 mm in breadth (Table 4 and Figure 2: A to D).

The scolex is small and globular, measuring 0.15 to 0.18 mm, with everted rostellum up to the base of suckers, and 0.12 to 0.14 mm without rostellum. It measures 0.22 to 0.23 mm across suckers. The rostellum with its sac measures 0.17 to 0.21 mm in length and extends below the posterior margin of suckers. The rostellum bears a single crown of 10 wrench-shaped hooks which measure 26 to 28 μ in length. The handle of hook is larger than the blade and is parallel to the equally long blade. The suckers are rounded and muscular, measuring 0.06 to 0.08 mm in diameter. The scolex is followed by a short neck that measures 0.17 to 0.22 mm in length and 0.10 to 0.15 mm in breadth. The dorsal longitudinal excretory duct is 10 to 13 μ and ventral excretory duct is 16 to 20 μ in diameter.

Testes are three in number, disposed in a triangle, two being aporal and one poral. They measure 92 to 100 μ in diameter. Cirrus pouch extends into the medullary

Table 4. Comparative measurements of *Mayhewia kavini* Chishti and Khan, 1982 with present form. (measurements in mm unless stated otherwise).

Particulars	Chishti and Khan, 1982	Present author
	<i>Mayhewia kavini</i>	
Max. length	60-90	55-84
Max. breadth	0.72-0.785	0.77-1.0
Mature proglottids	0.22-0.25 × 0.65-0.72	0.20-0.26 × 0.8-0.92
Gravid proglottids	0.26-0.35 × 0.72-0.785	0.3-0.36 × 0.77-1.0
Scolex length upto rostellum	0.19-0.198	0.15-0.18
Scolex length without rostellum	0.138-0.146	0.12-0.14
Width across suckers	0.242-0.249	0.22-0.23
Rosetellum length	0.222-0.228	0.17-0.21
Number of rostellar hooks	10	10
Hook size	25-30 μ	26-28 μ
Sucker size	0.09-0.1 × 0.075-0.082	0.06-0.08 (Dia.)
Dorsal excretory duct	10-12 μ	10-13 μ
Ventral excretory duct	18-22 μ	16-20 μ
Testis diameter	72-84 μ	93-100 μ
Cirrus pouch extent	Medullary	Medullary
Cirrus pouch size	0.14-0.155 × 0.052-0.057	0.16-0.19 × 0.050-0.053
External vesicula seminalis	0.16-0.18 × 0.06-0.072	0.18-0.25 × 0.07-0.072
Internal vesicula seminalis	0.06-0.072 × 0.048-0.052	0.05-0.065 × 0.42-0.50
Genital pore position	Anterior 1/3 rd.	Anterior 1/3 rd.
Ovary extend across	0.172-0.23 × 0.052-0.09	0.25-0.3 × 0.06-0.08
Vitelline gland	50-60 μ × 40-50 μ	65-72 μ × 48-53 μ
Receptaculum seminis	0.121-0.15 × 0.088-0.11	0.14-0.16 × 0.07-0.08
Egg size	42-54 μ × 30-41 μ	38-44 μ × 28-33 μ
Embryo size	28-32 μ × 18-22 μ	22-26 μ × 16-20 μ
Embryonic hook size	12-15 μ	9-12 μ

**Figure 1.** *Dilepis undula* Schrank, 1788. (A, B) Scolex showing suckers and rostellum with hooks; (C) rostellum showing hooks; (D, E) mature proglottids showing reproductive organs.

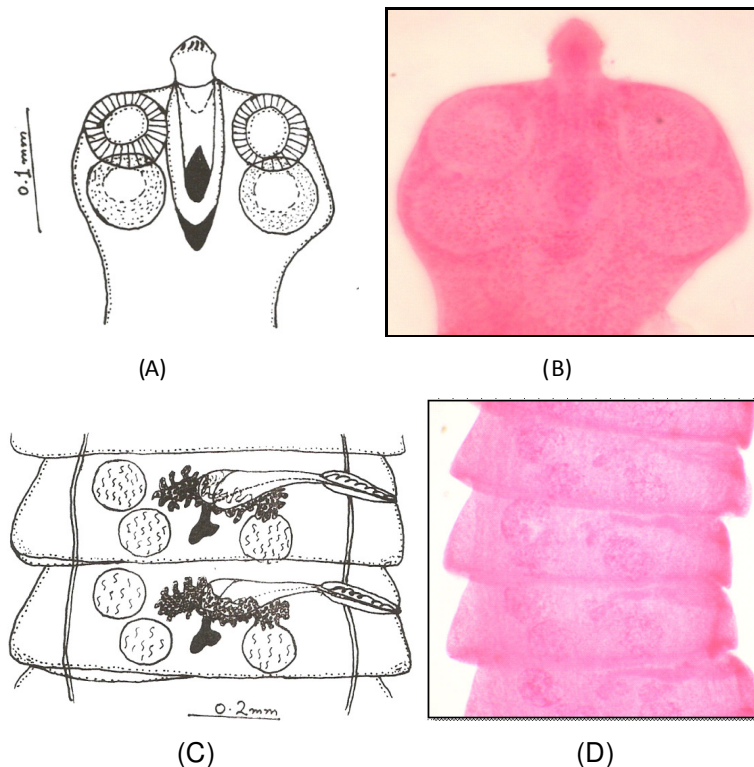


Figure 2. *Mayhewia Kavini* Chishti and Khan, 1982. (A, B) Scolex showing suckers and rostellum with hooks; (C, D) mature proglottids showing reproductive organs.

region and measures 0.16 to 0.19 mm in length and 0.05 to 0.053 mm in width. The external vesicula seminalis is large, extending to the middle of progottids and measures 0.18 to 0.25 mm in length and 0.07 to 0.072 mm in width. The small and oval internal vesicula seminalis measures 0.05 to 0.065 mm \times 0.042 to 0.05 mm. The genital pores are unilateral, opening in the anterior third margin of proglottid laterally. The follicular ovary lies in the middle of proglottid and is bilobed. It measures 0.25 to 0.3 mm across and 0.06 to 0.08 mm in thickness. Vitelline gland is slightly lobed and lies posterior to the ovary. It measures 65 to 72 \times 48 to 53 μ . The receptaculum-seminis is flask-shaped, situated anterior to ovary and measures 0.14 to 0.16 \times 0.07 to 0.08 mm. Receptaculum seminis narrows into a slender vaginal tube which opens into the genital atrium just posterior to the opening of cirrus pouch. The gravid proglottids are compactly filled with eggs which are oval in shape and measure 38 to 44 \times 28 to 33 μ . The embryo measures 22 to 26 \times 16 to 20 μ , and embryonic hooks measure 9 to 12 μ in length. In addition to *D. undula* and *M. kavini* redescribed in this paper, concurrent infestation with other helminth parasites such as *Echinostoma revolutum*, *Capillaria anatis*, *Anomotaenia galbulae* and *Choanotaenia micracantha* was also observed in some of the bird hosts during the present investigation.

DISCUSSION

The distinguishing characteristics of the present *Dilepis* specimens are: 46 to 58 rostellar hooks arranged in double crown; that of the anterior and posterior crown measure 74 to 85 and 67 to 78 μ , respectively; testes 24 to 32 in number, postovarian; cortical position of cirrus pouch; large bilobed ovary and unilateral genital pores.

When compared with the known species of genus *Dilepis* Weinland, 1858 described from different avian hosts, the present specimens shows a large similarity of characters with *D. undula* (Schrank, 1788). This species has also been redescribed several times, giving variation in the number and size of rostellar hooks, number of testes and length of strobila (Table 1). However, the present measurements come in the range of those described by various authors. In view of these measurements and other similarities as described above, the present cestode specimens are assigned to *D. undula* (Schrank, 1788). It is recorded for the first time in *Corvus* species from Kashmir. The earliest record of this species in Kashmir is from *Sturnus vulgaris humii* and *Turdus unicolor* (Chishti, 1974). The present cestode specimens of genus *Mayhewia* collected from *C. monedula* possess characteristics like rostellum with single crown of 10 small hooks; strobila comprising of numerous transversely

elongated progottids; two pair of excretory stems; three testes arranged in a triangle; cirrus pouch extending into medullary region; external and internal seminal vesicles present; ovary bilobed and uterus succular. All these characters are in conformity with the description of genus *Mayhewia* Yamuguti, 1956.

While comparing with the known species of genus *Mayhewia* Yamaguti, 1956, the present form shows a similarity of characters with those of *Mayhewia kavini* Chishti and Khan, 1982 as regards the number (10) and size of rostellar hooks, number of testes, extent of cirrus pouch, presence of seminal vesicles etc. However, some intraspecific variations were recorded in different structures as indicated in Table 2, the reason for which could be the age of parasite, host species, intensity of infection (higher intensity, smaller parasites), methodology (fixation), environmental factors of the study area, body conditions of the host, etc. (Ternopolskaya, 1984; Kuchai et al., 2012). Since these variations are of minor significance, therefore the present specimens in view of above similarities, are assigned to *Mayhewia kavini* Chishti and Khan, 1982.

Conclusion

The present paper redescribes only two species of cestode parasites (*D. undula* and *M. kavini*) recovered from three species of the genus *Corvus*. This work adds to the parasite species diversity infecting these avian hosts and may prove helpful for the future research on helminthes of birds. In addition, the present study showed that *M. kavini* was recovered only from *C. monedula*, hence the parasite may be host specific in nature.

ACKNOWLEDGEMENTS

The authors would like to thank all the members of the Department of Zoology, University of Kashmir who helped during the study. The authors express their deep sense of gratitude and sincere thanks to Prof. M. Z. Chishti, Professor Emeritus, Centre of Research for Development (CORD), University of Kashmir for helping in the identification of the cestode parasites.

REFERENCES

- Benesh DP (2010). Developmental inflexibility of larval tapeworms in response to resource variation. *Int J. Parasitol.* 40(4):487-497.
- Calnek BW (1997). *Diseases of Poultry*. 10th Ed. Iowa State University Press, Ames, Iowa 50014. pp. 850-851.
- Chishti MZ (1973). On a new species of the cestode genus *Choanotaenia* Railliet, 1896 from *Acridotheres tristis* in Kashmir. *J. Sci. Univ. Kashmir* 1(1-2):51-54.
- Chishti MZ (1980a). A new record of *Choanotaenia gondwana* Inamdar, 1934 (Choanotaeniidae: Cestoda) from *Passer domestica* in Kashmir. Proceedings of the 5th All India Congress on Zoology, Bhopal, India.
- Chishti MZ (1980b). *Dilepsis fotedari* n. sp. (Dilepididae Fuhrmann 1907: Cestoda) from *Anas platyrhynchos* in Kashmir. *Indian J. Helminthol.* 32(1):1-3.
- Chishti MZ (1980c). On the infection of Cestode genus *Sobolevicanthus* Spassky et Spaskeja, 1954 from Aves in Kashmir. Proceedings of the 3rd National Congress on Parasites, Hissar. pp. 13-14.
- Chishti MZ (1981). On a new species of the genus *Choanotaenia* Railliet, 1826 from *Corvus monedula* in Kashmir. Proceedings of the 68th Session of the Indian Science Congress.
- Chishti MZ, Khan AR (1978). Epidemiology of Cestode infection in Snipe, *Tringa hypoleuca* from Kashmir, with description of a new species of the genus *Amoebotaenia*. Proceedings of the 65th Session of the Indian Science Congress. p 316.
- Chishti MZ, Khan AR (1979). A new record of *Dilepsis undula* (Schrank, 1788) from some avian hosts in Kashmir. Proceedings of the 66th Session Indian Science Congress. p 107.
- Chishti MZ, Khan AR (1982). *Mayhewia kavini* sp. nov. (Hymenolepididae Railliet et Henry, 1909: Cestoda) from *Corvus monedula* in Kashmir. *Indian J. Helminthol.* 34(2):139-142.
- Chishti MZ, Mir AA, Rasool A (1986). *Choanotaenia micracantha* sp. nov. (Dilepoidea: Cestoda) from *Corvus monedula* in Kashmir. *Indian J. Helminthol.* 38(2):107-111.
- Dar GH, Bhagat RC, Khan MA (2002). Biodiversity of the Kashmir Himalaya, 1st Ed. Valley Book House, Kashmir University Road, Srinagar-190006, India.
- Davies TI (1935). The anatomy of *Dilepis undula* (Schrank, 1788). *Proc. Zool. Soc. London.* pp. 717-722.
- Fotedar DN, Chishti MZ (1973). On a new species, *Anomotaenia kashmirensis* (Choanotaeniidae, Methevossian 1953) from *Sturnus vulgaris* in Kashmir. *J. Sci. Univ. Kashmir* 1(1-2):48-50.
- Fotedar DN, Chishti MZ (1976). *Anomotaenia acrocephali* n. sp. and first record of *A. galbulae* (Gmelin, 1709) Fuhr; 1932 from some birds of Kashmir. *Riv. Parassitol.* 37(2):247-252.
- Fotedar DN, Chishti MZ (1976). *Pseudoschistotaenia* n. gen. (Amabillidae, Fuhrman, 1908: Cestoda) from *Podiceps ruficollis capensis* in Kashmir. Proceedings of 63rd Session Indian Science Congress, Part III (Abt). p 21
- Fotedar DN, Chishti MZ (1977). On a new species of the genus *Pseudoschistotaenia* Fotedar and Chishti, 1976. Proceedings, All Indian Symposium of Helminthology held at Srinagar (Abt). p 37.
- Fotedar DN, Mahajan, R, Dhar RL, Chishti MZ (1970). New variety of *Raillietina* from common Blue Rock Pigeon in Kashmir. *Kashmir Sci.* 7(1-2):103-106.
- Fotedar DN, Chishti MZ (1974). Redescription of *Choanotaenia oriole* Joyeux et Baer 1955 and *C. infundibulum* (Bloch, 1979) with a note on the synonymy of *C. dutii* Mukherjii 1964. *J. Sci. Univ. Kashmir* 2(1-2):73-78.
- Goodwin D (1983). *Crows of the World*. Queensland University Press, St Lucia, Qld.
- Gupta SP (1967). Helminthic-fauna of Kashmir. *Kashmir Sci.* 4(12):56-61.
- Henrich T, Daniel PB, Martin K (2013). Hybridization between two cestode species and its consequences for intermediate host range. *Parasit. Vectors* 6:33.
- Khan AR, Chishti MZ (1982). On *Echinostoma revolutum* (Froelich) Looss, 1899 and synonymy of *Neoechinostoma spinosa* Agarwal, 1963. *Kashmir Univ. Res. J.* 2:22-24.
- Kuchai JA, Fayaz A, Chishti MZ, Tak H, Javid AD, Dar SA, Muzaffar R (2012). A study on morphology and morphometry of *Haemonchus contortus*. *Pak. J. Zool.* 44(6):1737-1741.
- Kornyushin VV, Salamatin RV, Greben OB, Georgiev BB (2009). *Spiniglians sharpiloi* sp. n. (Cestoda, Dilepididae), a parasite of the common magpie, *Pica Pica*, in the palaeartic. *Vestn. zool.* 23:85-93.
- Lockie JD (1956). The Food and Feeding Behavior of the Jackdaw, Rook and Carrion Crow. *J. Anim. Ecol.* 25(2):421-428.
- Meyer CM, Olsen WO (1975). *Essentials of Parasitology*. WM. C. Brown Company Publishers, Dubuque, Iowa (USA).
- Mettrick DF (1958). Helminth parasites of Hertfordshire Birds-II Cestoda. *J. Helminthol.* 32:157-194.
- Nanware SS, Dhondge RM, Bhure DB (2011). Biosystematic studies on *Cotugnia orientalis* sp. nov. (Cestoda: Davaineidae, Fuhrmann 1907) from *Gallus gallus domesticus*. *Bioscan (Ranchi)* 6(1):71-75.
- Nyari A, Ryall C, Peterson AT (2006). Global invasive potential of the house crow *Corvus splendens* based on ecological niche modeling. *J. Avian Biol.* 37:306-311.

- Roberts LS, Janovy JJ (2005). Gerald D. Schmidt and Larry S. Roberts' Foundation of Parasitology. 7th Ed., Mc Graw-Hill Companies. p 311.
- Schmidt GD (1986). Handbook of Tapeworm Identification. CRC Press, Boca Raton, FL.
- Ternopolskaya LD (1984). Variability of *Fasciola hepatica* L., 1785 in different hosts. Bulletin Vsesoyuznogo Instituta Gel. Mintologii Im. K. I. Skrybin. 38: 47-51.
- Yamaguti S (1961). Systema Helminthum. Vol. 2. The Cestodes of Vertebrates. Interscience Publisher, John Wiley Sons, New York.
- Yoneva A, Céline L, Pavel NN, Yana M, Jean M, Boyko BG (2012). Spermiogenesis and spermatozoon ultrastructure of the paruterinid cestode *Notopentorchis* sp. (Cyclophyllidea). Parasitol. Res. 111(1):135-142.

UPCOMING CONFERENCES

24th International Conference of the World Association for the Advancement of Veterinary Parasitology, Perth, Australia, 25 Aug 2013



XXI Latin American Congress of Parasitology, Guayaquil, Ecuador, 6 Oct 2013



Conferences and Advert

August 2013

24th International Conference of the World Association for the Advancement of Veterinary Parasitology, Perth, Australia, 25 Aug 2013

September 2013

5th European Congress of Virology, Lyon, France, 11 Sep 2013

October 2013

XXI Latin American Congress of Parasitology, Guayaquil, Ecuador, 6 Oct 2013

Journal of Parasitology and Vector Biology



Related Journals Published by Academic Journals

- *Journal of Diabetes and Endocrinology*
- *Journal of Veterinary Medicine and Animal Health*
- *Research in Pharmaceutical Biotechnology*
- *Journal of Physiology and Pathophysiology*
- *Journal of Infectious Diseases and Immunity*
- *Journal of Public Health and Epidemiology*

academicJournals